



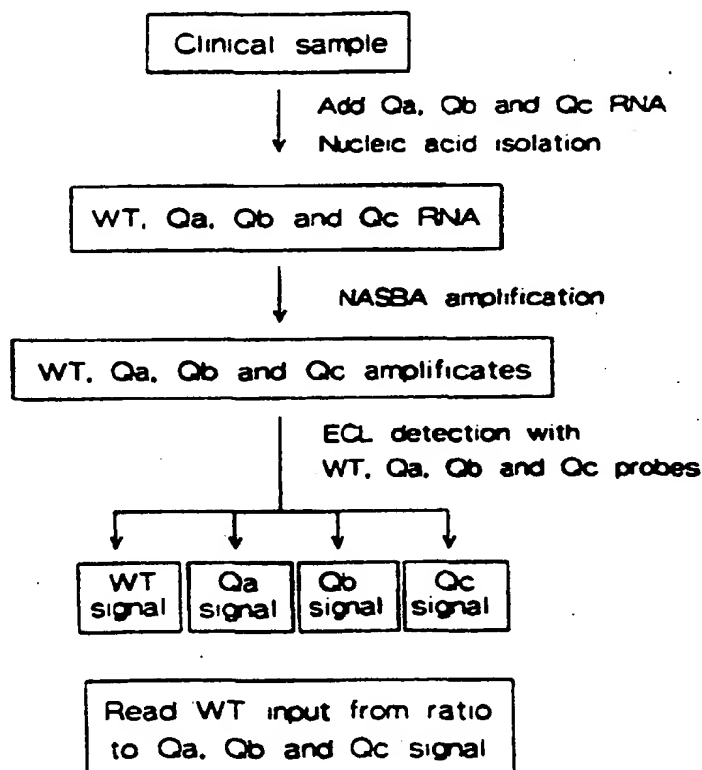
INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁶ : C12Q 1/68, 1/70	A1	(11) International Publication Number: WO 95/02067 (43) International Publication Date: 19 January 1995 (19.01.95)
(21) International Application Number: PCT/EP94/02295 (22) International Filing Date: 8 July 1994 (08.07.94) (30) Priority Data: 93202015.9 9 July 1993 (09.07.93) EP (34) Countries for which the regional or international application was filed: NL et al. 93203424.2 6 December 1993 (06.12.93) EP (34) Countries for which the regional or international application was filed: NL et al. (71) Applicant (for all designated States except US): AKZO NOBEL N.V. [NL/NL]; Velperweg 76, NL-6824 BM Arnhem (NL). (72) Inventors; and (75) Inventors/Applicants (for US only): BEUNINGEN, Marinus, Gerardus, Johannes [NL/NL]; Polderkade 27, NL-5345 RR Oss (NL). KIEVITS, Tim [NL/NL]; Van Miertstraat 20, NL-5262 XB Vught (NL). BEUMER, Thomas, Augustinus, Maria [NL/NL]; Granietstraat 63, NL-5345 SX Oss (NL). (74) Agent: HERMANS, F.G.M.; Postbus 20, NL-5340 BH Oss (NL).		(81) Designated States: AU, CA, FI, JP, KR, US, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE). Published <i>With international search report.</i> <i>Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i>

(54) Title: **IMPROVED METHOD FOR THE QUANTIFICATION OF NUCLEIC ACID**

(57) Abstract

The present invention is related to an improved method for the quantification of nucleic acid, which can be performed with a minimal amount of nucleic acid amplification reactions. The method according to the invention for the quantification of analyte nucleic acid in a sample comprises the steps of: adding to the sample different respective amounts of different nucleic acid constructs, each construct being distinguishable from the analyte nucleic acid and capable of being co-amplified with the analyte nucleic acid; subjecting the sample to a nucleic acid amplification procedure, using amplification reagents capable of reacting with both the analyte nucleic acid and the nucleic acid constructs; detecting the relative amounts of amplicates derived from analyte nucleic acid and each nucleic acid construct; calculating the amount of analyte nucleic acid from said relative amounts. Each nucleic acid construct is different; the nucleic acid constructs can be distinguished from one another and from the analyte nucleic acid. The nucleic acid constructs do resemble each other, and the analyte nucleic acid, in that all are capable of reacting with the same amplification reagents.



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Improved method for the quantification of nucleic acid.

The present invention is related to an improved method for the quantification of nucleic acid. The quantification of nucleic acid can be of great importance in various research as well as diagnostic areas. Quantification of nucleic acid can be an important tool in understanding gene regulation and may also be used in monitoring the effects of therapy. The amount of specific nucleic acid sequences present in a sample, as for example human blood or other bodily fluids, may also provide valuable information with respect to the disease state of persons infected with, for example, a certain virus and the efficiency of treatment with certain medicines.

Various methods for the quantitative amplification of nucleic acid have been described.

In WO 91/02817 a method for the quantification of nucleic acid using the Polymerase Chain Reaction (PCR) as amplification technique, is described. This method comprises the addition of a standard nucleic acid segment to a sample, which segment can react with the same primers that are used for the amplification of the unknown quantity of target nucleic acid present in the sample. Following amplification the amount of each of the two PCR products is measured, and the amount of the target segment in the original sample is quantified by extrapolating against a standard curve. The standard curve is generated by plotting the amount of the standard segment produced in a polymerase chain reaction against varying, but known, amounts of the nucleic acid present before amplification.

The addition of other sequences to a sample comprising analyte nucleic acid, which are capable of being coamplified with the analyte nucleic acid, has also been described by Becker and Hahlbrock (Nucleic Acids Research, Vol. 17, Number 22, 1989). The method described by Becker and Hahlbrock is a method for the quantification of

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nucleic acid where different known amounts of internal standard, comprising a nucleic acid sequence that differs from the analyte nucleic acid by just one nucleotide (point mutation) are added to fractions of known volume of a sample containing an unknown amount of analyte nucleic acid and coamplified by PCR with the analyte nucleic acid. Identical portions of total RNA are thus "spiked" with decreasing known amounts of internal standard RNA. By introducing one base change in the internal standard sequence a specific restriction site is created and the mutant sequence is cut with the appropriate restriction enzyme before the sample containing the amplified nucleic acid is applied to an electrophoretic gel. The nucleic acid is quantified by comparing the bands in the gel representing (a part of) the mutant sequence and the analyte nucleic acid. One of the disadvantages of this method is that incomplete digestion of the internal standard by the restriction enzyme might cause inaccuracies in the determination of the amount of nucleic acid present.

Becker and Hahlbrock use the internal standard as a marker in their quantification method where both analyte and marker are non-competitively amplified.

A method for the quantification of nucleic acid, using the addition of a known number of molecules of a nucleic acid sequence, corresponding to the target nucleic acid, to a sample containing an unknown amount of the target nucleic acid sequence has also been described in co-pending co-owned European patent application published under no. EP 525882. The method as described in EP 525882 is based on the principle of amplification of nucleic acid from a sample containing an unknown concentration of wild-type target nucleic acid, to which has been added a known amount of a well-defined mutant sequence. Amplification is performed with one primer set capable of hybridizing to the target as well as the mutant sequence. The competitive amplification described in EP 525882 can be performed with a fixed amount of sample and dilution series of mutant sequence or vice versa.

The above described methods involve the addition of a well defined standard sequence to the amplification mixture. To be able to obtain an accurate and reliable measurement with the above described method many amplification reactions have to be carried out: The sample has to be subdivided in different portions of known quantity to each of which a different, known, amount of standard nucleic acid has to be added. Or, with the method as described in WO 91/02817, a standard curve has to be generated from a dilution series, which is rather laborious and may also result in inaccurate estimations of the amount of nucleic acid present since the actual amplification concerning the target nucleic acid and the amplifications concerning the standard curve are performed separately.

Due to the great amount of amplification reactions that have to be carried out in order to be able to determine the amount of nucleic acid in a precise and reliable manner, the above described methods are very laborious and time consuming. The need therefore exists for a method for the quantification of nucleic acid that is less laborious and time consuming. The present invention provides such a method.

The present invention provides a method for the quantification of analyte nucleic acid in a sample comprising the steps of:

- adding to the sample different respective amounts of different nucleic acid constructs, each construct being distinguishable from the analyte nucleic acid and capable of being co-amplified with the analyte nucleic acid,
- subjecting the sample to a nucleic acid amplification procedure, using amplification reagents capable of reacting with both the analyte nucleic acid and the nucleic acid constructs,
- detecting the relative amounts of amplicates derived from the analyte nucleic acid and each nucleic acid construct,
- calculating the amount of analyte nucleic acid from said relative amounts.

With the method according to the present invention different nucleic acid constructs are added to the sample.

Each nucleic acid construct is different; the nucleic acid constructs can be distinguished from one another and from the analyte nucleic acid. The nucleic acid constructs do resemble each other, and the analyte nucleic acid, in that all are capable of reacting with the same amplification reagents. With amplification reagents, among other things, one or more amplification primers are meant, which are capable of hybridizing to the analyte nucleic acid and the nucleic acid constructs. Other amplification reagents are the usual reagents used with amplification procedures like the necessary enzymes, nucleic acid polymerases, used with the different amplification techniques known in the art. The method according to the invention can be used with any kind of amplification procedure, for example the so-called polymerase chain reaction (PCR) as described in USP 4,683,195 and 4,683,202. Another method for the amplification of nucleic acid is the "nucleic acid sequence based amplification (NASBA)" as disclosed in European Patent application EP 0,329,822.

The nucleic acid constructs used with the method according to the present invention are nucleic acid sequences that resemble the analyte nucleic acid in that they are capable of reacting with the same amplification reagents. Each nucleic acid construct should therefore at least comprise the sequence to which nucleic acid amplification primers used can anneal, which sequence is also present in the analyte nucleic acid. The nucleic acid constructs used with the method according to the invention can be distinguished from the analyte nucleic acid and from each other. This can be achieved by constructing nucleic acid constructs in a such a way that each construct comprises a uniquely distinguishable sequence that is not present in the other nucleic acid constructs used nor in the analyte nucleic acid. Preferably nucleic acid constructs are used

wherein the uniquely distinguishable sequence is a randomized sequence, comprising e.g. about 20 nucleotides, thus keeping the length and nucleotide constitution of constructs and analyte nucleic acid the same. In the detection, following amplification of analyte nucleic acid and nucleic acid constructs present in a sample, the different nucleic acid constructs may be measured separately by using different detection probes, each capable of recognizing only one of the unique sequences present in the nucleic acid constructs.

Of course there are other ways in which the nucleic acid constructs can be constructed that will render them unique. However, preferably the nucleic acid constructs should not be mutated in such a way that the amplification is hampered. Preferably, with the method according to the invention, nucleic acid constructs are used that are capable of being amplified with the same efficiency as the analyte nucleic acid, otherwise it is difficult to make an accurate calculation based on the amounts of analyte and construct nucleic acid present in the sample after amplification.

Nucleic acid constructs are polynucleotides that can be prepared by different methods known in the art. Nucleic acid constructs can, for example, be prepared by various recombinant DNA strategies. For instance, new sequences can be introduced in the analyte sequence by digestion of the plasmid containing the analyte sequence with restriction enzymes, thus removing the original sequence and inserting a new sequence that may be prepared on a nucleic acid synthesizer or subcloned from a different plasmid. Complete nucleic acid constructs may also be prepared, using a nucleic acid synthesizer.

The nucleic acid constructs should be added to a sample, containing an unknown amount of analyte nucleic acid, prior to subjecting the sample to an amplification procedure. The sample, to which a known quantity of different nucleic acid constructs are added, should, of

course, have a predetermined volume, in order to be able to calculate the concentration of nucleic acid present in the sample later on. The sample has to contain a known volume, or quantity of material, taken from the material in which the amount of nucleic acid should be determined. When the concentration of nucleic acid in a certain test fluid is to be determined, a sample should contain all nucleic acid isolated from a known quantity of the test-material under investigation.

The nucleic acid constructs may be added prior or after subjecting the test fluid under investigation (for example human blood serum) to a procedure for the isolation of nucleic acid. Methods for the isolation of nucleic acid have been described and are known to anyone skilled in the art. Nucleic acid isolation procedures, that may be used to prepare a sample that is to be subjected to the method according to the present invention, have been described in, for example, Maniatis et al., Molecular Cloning, a laboratory manual, Cold Spring Harbor Laboratory. Another method for the processing of nucleic acid containing samples has been described by Boom et al., J.Clin.Microbiol. 28:495-503, 1990.

In a preferred embodiment of the invention the nucleic acid constructs are added prior to isolating the nucleic acids from the quantity of test fluid. In this way the loss of nucleic acid that might optionally occur during the isolation procedure will be reflected in both the resulting amounts of analyte nucleic acid and construct nucleic acid present in the sample. The amount of analyte nucleic acid present in original test fluid under investigation can be calculated directly from the obtained results.

When the nucleic acid constructs are added to a known quantity of test fluid prior to subjecting the test-fluid to a nucleic acid isolation procedure, an isolation control nucleic acid construct can be added to the sample after

the nucleic acid isolation procedure has been performed. With the addition of the IC (isolation control) sequence it is possible to determine the efficiency of the nucleic acid isolation procedure. Thus, for each sample the threshold value, can be adjusted to the isolation efficiency. The isolation control construct can be a sequence that differs from the analyte and the other nucleic acid constructs, for example because it comprises a unique sequence, but will preferably be amplified with the same efficiency. The IC can be regarded as another "nucleic acid construct" the difference being that the IC molecules are added after the nucleic acid isolation procedure is performed, while, in this preferred embodiment of the invention, the nucleic acid constructs are added prior to subjecting the test fluid to a nucleic acid isolation procedure. The efficiency of the isolation procedure can be calculated, since a well defined amount of molecules of each nucleic acid construct was added prior to subjecting the test-fluid to a nucleic acid isolation procedure, and a well defined amount of IC molecules was added after the nucleic acid isolation procedure was performed. When, for example, 100 molecules of a particular nucleic acid construct were added and the same amount of IC molecules was added, a 100% isolation efficiency would result in the same signal (after amplification) for both the nucleic acid construct and the IC. If the IC signal obtained would be twice as high as the signal obtained for the nucleic acid construct the efficiency of the nucleic acid isolation procedure was only 50% (meaning that half of the nucleic acid molecules originally present were lost in the isolation procedure). The threshold value can now be adjusted accordingly, thus lowering the risk of obtaining false-negative test results.

More than one nucleic acid construct is added, each nucleic acid construct in a different known quantity. Preferably the nucleic acid constructs are added in a range of amounts, differing from each other by a constant

factor (e.g. a factor 10). For example, when three nucleic acid constructs are used, QA, QB, and QC, 10^2 molecules of QA 10^3 molecules of QB and 10^4 molecules of QC, may be added to the sample.

The sample may also be subdivided in more than one reaction aliquots, each of a known volume, to each of which nucleic acid constructs are added in different ranges of amounts. Preferably, in that case, to each reaction volume the same nucleic acid constructs are added. If, for example, two reactions are used, a "low-range" reaction and a "high range" reaction can be carried out. For example, when the ranges of nucleic acid constructs would overlap, to the low-range reaction volume QA, QB, and QC may be added in the quantities as mentioned before, while to the high-range reaction volume 10^4 10^5 and 10^6 molecules of, QA, QB and QC are added respectively. The ranges of amounts of nucleic acid constructs may overlap but there can also be a gap between the amounts of nucleic acid constructs in the low range and in the high range respectively. Although the use of more than one reaction volume means that more than one amplification reaction has to be carried out, the saving of work and time compared to the prior art methods for the quantification of nucleic acid is still substantial, since with a minimum of reactions a very broad range of analyte concentrations may be covered.

The advantage of using ranges of amounts of nucleic acid constructs that do not overlap is that the number of constructs used may be reduced, while still a broad range of concentrations can be covered. When the analyte is present in an amount that falls in the gap between the ranges of amounts of nucleic acid constructs, no direct comparison to a signal range originating from nucleic acid constructs present in the sample or reaction aliquot, will be possible. However, the amount of analyte nucleic acid can still be determined since the signal of the analyte nucleic acid will be stronger than the low-range signals and weaker than the high range signals, and the concentration can be calculated with these data.

When, with the method according to the invention different ranges of nucleic acid constructs are added to more than one reaction volume, the sample in which the concentration of analyte nucleic acid is to be determined can be split into different volumes, and the nucleic acid constructs can be added to these volumes, before applying each volume to a nucleic acid isolation procedure. Although each volume will have to be treated separately in an isolation procedure, no corrections have to be made for optional losses of analyte nucleic acid during isolation, since variations in performance are corrected for by the addition of the nucleic acid constructs that undergo exactly the same treatment and suffer from the same variations. Of course, one may also subject the total volume to a nucleic acid isolation procedure and split it into different volumes, to which the ranges of constructs will then be added, afterwards.

A problem that may arise when performing amplification reactions is that samples may get contaminated with nucleic acid molecules originating from other samples that, for example, were tested previously in the same laboratory. This may result in false positive test results or, in the case of quantitative measurements, a false outcome in the calculation of the number of nucleic acid molecules present in the sample.

Contaminations, of samples tested with the method according to the present invention, in the range of 10-100 molecules will not alter the internal calibration line and will only effect the calculated amount of molecules of analyte nucleic acid if the contamination consists largely of analyte molecules and the amount of analyte nucleic acid in the sample being tested is less than 100 molecules. Contaminations with about 100-1000 molecules will slightly influence the internal calibration line and will only interfere with the calculation of the amount of analyte nucleic acid molecules present in the sample if the amount of analyte molecules in the sample is less than

1000 molecules. Contaminations with more than 1000 molecules will alter the calibration line and the outcome of the calculation.

With a preferred embodiment of the present invention the contamination of samples with nucleic acid molecules originating from previously tested or other samples can be detected.

Contamination can be detected if, when the method according to the invention is used to test more than one sample, the same nucleic acid constructs are used in all samples and the amounts of each particular nucleic acid construct are varied. For example, when three nucleic acid constructs are used (Q_a , Q_b and Q_c), to sample "A" these constructs may be added in the following amounts: $Q_{a_{high}}$, $Q_{b_{middle}}$ and $Q_{c_{low}}$. To sample "B" the same constructs may now be added in the amounts $Q_{a_{low}}$, $Q_{b_{high}}$, $Q_{c_{middle}}$, while to samples "C" the following amounts are added: $Q_{a_{middle}}$, $Q_{b_{low}}$, $Q_{c_{high}}$, etc. Of course the same formulation of nucleic acid constructs may also be used for more than one sample, for example 10 samples in row, before switching to the use of another formulation.

Up to six variations can be made, when three constructs are used, before the same formulation of nucleic acid constructs has to be used again. When four constructs are used up to twenty-four variations can be made etc. The use of different formulations of nucleic acid constructs in different samples will make a contamination, with nucleic acid material originating from a sample for which a different formulation was used detectable. The contamination will alter the internal calibration line in the sample under investigation; the calibration line will have another slope and a low correlation coefficient compared to normal data and will therefore be distinguishable as a contamination.

If, in each sample, a zero level of one of the constructs is used (and the construct for which a zero level is used varies per sample) even very low amounts of contaminating molecules can be detected. It is evident that the presence of a detectable amount of a particular construct of which

a zero amount was added, indicates that this particular sample was contaminated with material from a sample for which a different formulation had been chosen.

The detection of the nucleic acid constructs and the analyte nucleic acid after amplification can be performed in different ways. Many detection procedures that are known in the art can be used, as long as a measurable signal is generated that differs according to the amount of amplified nucleic acid (construct or analyte) which the signal represents. Detection methods that can be used with the method according to the invention include methods based on enzymatic and luminescent (fluorescent, electrochemiluminescent, fosforescent) phenomena as well as detection methods using solid labels like, for example, gold or dye sols and detection methods based on agglutination.

The concentration of analyte nucleic acid present in the sample can be calculated from the relative amounts of analyte nucleic acid and nucleic acid constructs that are represented by the signals generated during the detection procedure.

For example, the logarithm of the ratio between the signals representing the construct (Q) and analyte (A) sequences, $\log(Q/A)$, is a linear function of the logarithm of the amount of construct added to the sample, $\log(Q\text{-input})$. When the $\log(Q/A)$ is plotted as a function of $Q\text{(input)}$ a linear graph should be obtained, where the amount of analyte nucleic acid can easily be obtained from the intersection of the straight line with the X-axis. (Provided that the amount of analyte nucleic acid lies near the range of amounts of nucleic acid constructs added.)

Another embodiment of the present invention comprises a method for the quantification of analyte nucleic acid in a panel of test fluids, comprising the dilution of a quantity of said test fluids by a known factor. Samples

can now be taken from said diluted test fluids containing an amount of analyte nucleic acid expected to be within a certain range. When the amount of analyte nucleic acid in each one of a panel of test-fluids is to be determined, the complete panel can be pre-diluted to a concentration that is expected to be within this range. The pre-dilution should be carried out in such a way that most of the samples will fall within said range, while the remaining samples will all have a concentration below or will all have a concentration above said range. After the dilution step the complete panel may be subjected to the method according to the invention as described above, wherein the amounts of nucleic acid constructs added to the samples are within said range.

Remaining test-fluids taken from the panel that are below said range (or above) can be tested again with another adjusted dilution or undiluted samples. The advantage of this method is that less amplification reactions are needed than with the previous described high/low range separation.

In another embodiment of the method for the quantification of analyte nucleic acid in a sample according to the invention to a sample a quantity of a nucleic acid construct may be added, said quantity expected to be within the same range as the expected amount of analyte nucleic acid present in said sample, subjecting the sample to a nucleic acid amplification procedure, using amplification reagents capable of reacting with both the analyte nucleic acid and the nucleic acid construct, detecting the relative amount of amplicates derived from analyte nucleic acid and the nucleic acid construct and estimating the amount of analyte nucleic acid present in said sample from said relative amount. After this estimation has been made a second sample from the same test fluid can be subjected to the method according to the invention as described above wherein the amounts of nucleic acid constructs added are within the same range as the estimated amount of analyte nucleic acid present in the first sample.

The advantage of this procedure is that the range of amounts of constructs used can be chosen rather narrow, which will increase the accuracy of the method, and will lower the amount of amplification reactions that has to be carried out even further.

DESCRIPTION OF THE FIGURES:

Figure 1A: Input and amplified amounts of both nucleic acid constructs and sample (analyte) nucleic acid for experiment 1 as described in Example 1.

Figure 1B: Ratio of construct-signal and sample-signal plotted on a logarithmic scale against the amount of input construct copies as used in experiment 1 described in Example 1. The amount of analyte nucleic acid is indicated on the x-axis by the vertical line in the graph.

Figure 2A: Input and amplified amounts of both nucleic acid constructs and sample (analyte) nucleic acid for experiment 2 as described in Example 1.

Figure 2B: Ratio of construct-signal and sample-signal plotted on a logarithmic scale against the amount of input construct copies as used in experiment 2 described in Example 1. The amount of analyte nucleic acid is indicated on the x-axis by the vertical line in the graph.

Figure 3A: Input and amplified amounts of both nucleic acid constructs and sample (analyte) nucleic acid for experiment 3 as described in Example 1.

Figure 3B: Ratio of construct-signal and sample-signal plotted on a logarithmic scale against the amount of input construct copies as used in experiment 3 described in Example 1. The amount of analyte nucleic acid is indicated on the x-axis by the vertical line in the graph.

Figure 4A: Input and amplified amounts of both nucleic acid constructs and sample (analyte) nucleic acid for experiment 4 as described in Example 1.

Figure 4B: Ratio of construct-signal and sample-signal

plotted on a logarithmic scale against the amount of input construct copies as used in experiment 4 described in Example 1. The amount of analyte nucleic acid is indicated on the x-axis by the vertical line in the graph.

Figure 5A: Input and amplified amounts of both nucleic acid constructs and sample (analyte) nucleic acid for experiment 5 as described in Example 1.

Figure 5B: Ratio of construct-signal and sample-signal plotted on a logarithmic scale against the amount of input construct copies as used in experiment 5 described in Example 1. The amount of analyte nucleic acid is indicated on the x-axis by the vertical line in the graph.

Figure 6: Dynamic range of the one-tube Q-NASBA. For input of quantification $10^{1.44}$, $10^{2.83}$, $10^{4.23}$ and $10^{4.83}$ *in vitro* generated WT-RNA molecules were used. The quantification of every WT-RNA amount was performed 8-10 times. Q_A , Q_B and Q_C RNA were used at amounts of 10^4 , 10^3 and 10^2 RNA molecules, respectively

Figure 7: Schematic flow-chart of the one-tube Q-NASBA with addition of Q_A , Q_B and Q_C internal standard RNAs before nucleic acid isolation in the lysis buffer.

The present invention is further exemplified by the following examples:

EXAMPLES:

EXAMPLE 1:

The following are theoretical examples of a quantitative assay and subsequent detection performed according to the method of the present invention. From these experiments it can be seen how an assay according to the present invention may be performed, as well as the results that would be generated.

Five experiments that may be performed with the method according to the present invention are outlined below. The experiments differ in the amount of input analyte nucleic acid molecules and the ranges of nucleic acid constructs added prior to amplification.

The amounts of input analyte and input constructs are indicated in the table (table 1) given below:

Table 1: Amounts of input analyte and construct nucleic acid for experiment 1-5.

	Analyte Input	Low range Q Input	High range Q Input
Experiment 1:	150	QA: 100 QB: 1000 QC: 10000	QA: 10000 QB: 100000 QC: 1000000
Experiment 2:	2000	QA: 100 QB: 1000 QC: 10000	QA: 10000 QB: 100000 QC: 1000000
Experiment 3:	300.000	QA: 100 QB: 1000 QC: 10000	QA: 10000 QB: 100000 QC: 1000000
Experiment 4:	5000	QA: 100 QB: 1000	QA: 10000 QB: 100000
Experiment 5:	5000	QA: 100 QB: 1000	QA: 100000 QB: 1000000

As can be seen from this table, in the first three theoretical experiments three constructs are added (QA, QB and QC, respectively) and two amplification reactions are carried out with a "low range" of construct amounts and a "high range" of construct amounts. In these experiments the low range and high range do overlap. The overlapping point in these two ranges can be used to compare the amplifications. (If the amplifications are performed in the same way the signals generated should be the same for the overlapping amount of construct in the two ranges). In experiment four and five two ranges of amounts of constructs are used as well, but the low range and the high range do not overlap. Consequently the amount of constructs used per amplification reaction may be reduced, while the same total range of amounts will still be

covered.

The amplification rate of the Q-RNAs (constructs) and Wild-type RNA is the same since the RNAs are of the same size, differing only in a sequence of 20 randomized nucleotides, and the same primers and enzymes are used for the amplification. Therefore the initial ratio of the amount of each construct RNA and the amount of Wild-type RNA will not change during amplification. In the detection procedure following the amplification each amplificate mixture is split into four assays for the detection of Wild-type, QA, QB or QC RNA-amplificate. (The amplificate mixtures of experiment four and five only need to be split in three assays.)

If the log ratio of the construct signal and wild type signal is expressed against the logarithm of the input amount of construct RNA, a straight line is expected. The amount of wild type RNA can be calculated from this line. The results that can be obtained with the above described experiments are depicted in figure 1 to 5.

In the first graph of each figure, (a), the amounts of the different constructs, prior and after amplification, are presented, while the amount of analyte nucleic acid is indicated as well.

The second graph of each figure, (b), shows a graph wherein the log ratio of the expected construct signals and the analyte signal (sample signal) is expressed as a function of logarithm of the input amounts of constructs. The amount of analyte input nucleic acid molecules can be derived from these graphs, and follows from the logarithm of the input value (as depicted on the horizontal axis) belonging to the point on the vertical axis where the ratio is equal to 1.

EXAMPLE 2:

A mixture of quantified amounts of construct HIV-1 gag1 RNA transcript was made, comprising respectively 200, 2000 and 20000 copies of three nucleic acid constructs, QA, QB and QC. Quantified amounts of five different wild-type HIV-1 gag1 transcripts, respectively 200000, 20000, 2000, 200, 20 copies and a blank, were mixed with the mixture of Q construct RNAs.

Six amplification reactions, one for each amount of wt-RNA were performed according to a standard protocol. (T. Kievits et al).

From each amplificate 5 μ l was diluted to 100 μ l with a TBE buffer (90 mM Tris-Borate, 1 mM EDTA pH 8.4). From each dilution 5 μ l was added to a tube containing 15 μ l of a mixture of 3 pmol biotin-oligo, for capturing of all amplificates, 3 pmol of different tris(2,2'-bipyridine) Ruthenium II chelate labelled oligonucleotides, each comprising a specific sequence for either the wild-type RNA amplificates or one of the construct (QA, QB or QC) amplificates and 20 μ g streptavidin coated magnetic dynal beads M280 in a 6.67x SSC buffer (0.75 M NaCl, 0.075 M Sodiumcitrate pH 7-8).

Four hybridization assays for WT, QA, QB or QC detection were performed on one amplificate. The mixtures were hybridized for 30 minutes at 41°C and mixed every 10 minutes. 300 μ l of assay buffer for electrochemiluminescent ECL detection using the ORIGEN 1.5 detection system of IGEN was added and mixed.

The actual detection in the ORIGEN 1.5 detection system was performed according to manufacturers protocol.

EXAMPLE 3:

In this example a Q-NASBA assay in which 3 Q-RNA internal standards were spiked into the WT-RNA sample at amounts of 10^4 , 10^3 , 10^2 molecules is described. The 3 internal standard RNA molecules were distinguished using specific

ECL labelled probes for a 20 nucleotide randomized sequence (Van Gemen et al. J.Virol.Methods. 43, 177-188, 1993), specific for each internal standard. The ratio's of NASBA amplified internal standards and WT-RNA were measured using an ECL detection instrument.

ECL is based on chemiluminescent labels that emit light on the surface of an electrode (Blackburn et al., Clin.Chem. 37, 1534-1539, 1991). Detection of the signal can be quantified with a dynamic range over 5 orders of magnitude using a specifically developed detection instrument. The ECL technology has been adapted for the detection of amplified nucleic acid using ECL-labelled oligonucleotides in hybridization assays (Kenten, J.H. et al. Clin.Chem. 38, 873-879, 1992). Since the specific activities of the WT, Q_A , Q_B and Q_C ECL probes are known, the ratio's of WT, Q_A , Q_B and Q_C NASBA amplified RNA could be determined from the signal ratios of the respective probes.

The initial amount of WT input RNA was read from the ratio of the WT signal to Q_A , Q_B and Q_C signals in the ECL bead-based assay.

The way in which the Q-constructs were prepared and the assay was performed is described in the "materials and methods" section of this example.

The one-tube Q-NASBA protocol with ECL detection was used for quantifying different amounts of *in vitro* generated WT-RNA. Three different assays were performed using $10^{1.44}$, $10^{2.83}$, $10^{4.23}$ and $10^{4.83}$ initial WT-RNA molecules as input respectively. The Q-NASBA using the ECL bead-based assay was performed 8-10 times for each WT-RNA amount quantitated by mixing the WT-RNA with 10^4 Q_A RNA molecules, 10^3 Q_B RNA molecules and 10^2 Q_C RNA molecules in a single NASBA amplification. The results (mean \pm SD) of the quantifications performed were $10^{1.54 \pm 0.23}$, $10^{2.68 \pm 0.21}$, $10^{4.16 \pm 0.20}$ and $10^{4.81 \pm 0.23}$ using $10^{1.44}$, $10^{2.83}$, $10^{4.23}$ and $10^{4.83}$ initial WT-RNA molecules as input, respectively.

The results of the assays performed are depicted in figure 6. As can be seen from this figure the WT-RNA amount can be quantitated reliably to $1/10$ of the lowest Q-RNA amount used in the one tube quantification protocol.

MATERIALS AND METHODS

Plasmids and RNA synthesis

Genetic and recombinant DNA techniques followed standard procedures (Sambrook, J. et al., Molecular cloning. A laboratory manual, Cold Spring Harbor, NY: Cold Spring Harbor Lab., 1989. Ed. 2nd. The plasmid pGEM3RAN (Van Gemen et al. 1993) with a 22 nucleotide randomized sequence (pos. 1429-1451 HIV-1 pv22 sequence, (Muesing, M.A. et al., Nature 313, 450-458, 1985) was used to delete the Acc I site in multiple cloning site and part of the HIV-1 cloned sequence (pos 1691 to 2105 HIV-1 pv22) for reasons not related to the quantitative NASBA amplification. In this plasmid the randomized sequence was replaced by 5'ATG.CAA.GGT.CGC.ATA.TGA.GTA.A3' or 5'ATA.AGC.ACG.TGA.CTG.AGT.ATG.A3' to create pGEM3Q_BΔgag3 and pGEMQ_CΔgag3 respectively. Plasmid pGEM3RAN was renamed pGEM3Q_A. *In vitro* RNA was generated from pGEM3p24 (WT-RNA), pGEM3Q_A (Q_A-RNA), pGEM3Q_BΔgag3 (Q_B-RNA) and pGEM3Q_CΔgag3 (Q_C-RNA) using SP6 RNA polymerase (Sambrook, 1989). The cloned inserts of pGEM3p24 and pGEM3Q_A were recloned into vector pGEM4 creating pGEM4p24 and pGEM4Q_A. Using these plasmids *in vitro* RNA was made using T7 RNA polymerase (Sambrook, 1989). The length of the *in vitro* RNA is 1514 nucleotides for plasmids pGEM3p24 (WT-RNA) and pGEM3Q_A (Q_A-RNA) and 1090 nucleotides for plasmids pGEM3Q_BΔgag3 (Q_B-RNA) and pGEMQ_CΔgag3 (Q_C-RNA). The RNA was treated with DNase to remove plasmid DNA and purified on an anionic exchange column (Qiagen). The *in vitro* RNA was quantitated spectrophotometrically and diluted to desired concentrations with water. All RNA solutions were stored at -20°C.

Nucleic acid isolation

Nucleic acids were isolated from plasma according to the method of Boom et al. (Boom, R., et al., J.Clin.Microbiol. 28, 495-503, 1990; Van Gemen et al.1993). Nucleic acid of 100 μ l plasma was finally resuspended in 100 μ l water and stored at -70°C .

NASBA

All enzymes were purchased from Pharmacia, except AMV-reverse transcriptase which purchased from Seikagaku. BSA was purchased from Boehringer Mannheim. Twenty-three μ l NASBA reaction mixtures (final concentration in 25 μ l reaction mixture: 40 mM tris, pH8.5, 12 mM MgCl_2 , 42 mM KCl, 15% V/V DMSO, 1 mM each dNTP, 2 mM each NTP, 0.2 μ M Primer 1: 5' AAT.TCT.AAT.ACG.ACT.CAC.TAT.AGG.GTG. CTA.TGT.CAC.TTC.CCC.TTG.GTT.CTC.TCA, 0.2 μ M primer 2: 5' AGT.GGG. GGG.ACA.TCA.AGC.AGC.CAT.GCA.AA, 0.2-2 μ l wild-type RNA and 2 μ l *in vitro* Q-RNA; (Kievits, T. et al. J.Virol.Methods. 35, 273-286, 1991; Van Gemen et al.1993) were incubated at 65°C for 5 minutes to allow destabilization of secondary structures in the RNA and subsequently cooled down to 41°C to allow primer annealing. The amplification was started by adding 2 μ l enzyme mixture (0.1 $\mu\text{g}/\mu\text{l}$ BSA, 0.1 Units RNase H, 40 Units T7 RNA polymerase and 8 Units AMV-reverse transcriptase). Reactions were incubated at 41°C for 90 minutes. For every quantitation 2 negative controls were added.

Enzymatic bead-based detection

To detect and determine the ratio of NASBA amplified WT and Q_A RNA in the earlier described quantification protocol (Van Gemen et al.1993) a bead-based enzymatic assay was developed. One hundred μ l of 2.8 μ m polystyrene paramagnetic beads (Dynal Inc., Great Neck, N.Y., USA) coated with streptavidin were washed twice with 200 μ l 1xPBS, 0.1% BSA and resuspended in 100 μ l 1xPBS, 0.1% BSA. The washed beads were incubated 1 hr at room temperature with 300 pmol of a HIV-1 specific, biotinylated capture probe (5' TGT.TAA.AAG.AGA.CCA.TCA.ATG. AGG.A) and

subsequently washed once with 200 μ l 5xSSPE, 0.1% SDS and once with 200 μ l 1xPBS, 0.1% BSA. The beads were resuspended in 100 μ l, 1xPBS, 0.1% BSA.

Five μ l beads, 5 μ l NASBA reaction mixture and 50 μ l hybridization buffer (5xSSPE, 0.1% SDS, 0.1% blocking reagent, 10 μ g/ml Salmon sperm DNA) were incubated for 30 minutes at 45°C. The beads were washed twice with 100 μ l 2xSSC, 0.1% BSA followed by an incubation with 5×10^{-7} μ mol WT or Q detection oligonucleotide probe, of which 10% was HRP labelled, in 50 μ l hybridization buffer for 30 minutes at 45°C.

The bead-capture oligonucleotide-NASBA amplified WT or Q_A RNA-detection probe complex was washed once with 100 μ l 2xSSC, 0.1% BSA, once with 100 μ l TBST and twice with 100 μ l TBS. Subsequently, 100 μ l colour substrate (TMB/peroxide solution) was added to the beads and incubated for 3 minutes at room temperature.

The colour reaction was stopped by addition of 50 μ l 250 mM oxalate. The absorbance of the 150 μ l colour reaction was read at 450 nm in a micro-plate reader (Micro SLT 510, Organon Teknika, Boxtel, The Netherlands).

The absorbance values were corrected for background signal (i.e. negative controls) and the signal calculated as the percentage of the signal obtained by independently amplified WT or Q_A RNA.

ECL bead-based detection

Five μ l of NASBA amplified RNA (WT, Q_A, Q_B and Q_C) diluted 20 times in water was incubated with 20 μ l (3.3 pmol) of the HIV-1 specific, biotinylated capture probe (see enzymatic bead based detection), 3.3 pmol of an ECL (tris [2,2-bipyridine] ruthenium [II] complex) labelled oligonucleotide probe, specific for either WT, Q_A, Q_B or Q_C NASBA amplified RNA and 20 μ g (2 μ l) of streptavidin coated magnetic beads in 5 x SSC for 30 minutes at 41°C. During the incubation the tubes were mixed every 10 minutes by vortexing. Subsequently, 300 μ l of TPA solution (100 mM tripropylamine, pH=7.5) was added and the hybridization mixture was placed in an Origen 1.5 ECL

detection instrument (Organon Teknika, Boxtel, The Netherlands).

EXAMPLE 4:

The efficiency of the nucleic acid isolation can influence the outcome of the WT-RNA quantification. To circumvent the influence of loss of nucleic acid during the isolation, the Q_A , Q_B and Q_C RNA's can be added before or during the first step of the nucleic acid isolation (figure 7). This principle was tested using 10^4 molecules *in vitro* generated WT-RNA and HIV-1 RNA isolated from an *in vitro* cultured viral stock solution. *In vitro* generated WT-RNA and the HIV-1 viral stock RNA were re-isolated with and without the addition of Q_A , Q_B and Q_C RNA during the first step of the nucleic acid isolation (i.e. in the lysis buffer).

The methods used are described in the "materials and methods" section of example 3.

When originally isolated WT-RNA and HIV-1 viral stock RNA were quantified by the addition of Q_A , Q_B and Q_C during the amplification (i.e. control quantifications) the outcome was 3.3×10^4 and 2.1×10^4 for *in vitro* generated WT-RNA and HIV-1 viral stock RNA respectively. Quantification of the re-isolated RNA with addition of Q_A , Q_B and Q_C RNA before the re-isolation procedure revealed 2.5×10^4 and 1.5×10^4 for *in vitro* generated WT-RNA and HIV-1 viral stock RNA respectively. However, when the re-isolated RNA was quantified with addition of Q_A , Q_B and Q_C RNA after the re-isolation procedure the outcome was 4.7×10^3 and 2.0×10^3 for *in vitro* generated WT-RNA and HIV-1 viral stock RNA respectively. This indicates that the efficiency of re-isolation of RNA was approximately 10%. However, addition of Q_A , Q_B and Q_C before the re-isolation resulted in RNA quantifications that were equal to the control quantifications due to a constant ratio of WT: Q_A : Q_B : Q_C RNA, independant of the absolute amount of nucleic acid

isolated.

Results are depicted in Table 2.

Table 2: Comparison of re-isolation of 10^4 *in vitro* generated WT-RNA molecules and an HIV-1 viral stock RNA solution with addition of Q_A , Q_B , Q_C RNA during the first step of nucleic acid isolation and after the nucleic acid isolation .

Additions during NA isolation	Additions ^a after NA isolation	WT-RNA quantitated
	10^4 WT-RNA + Q_A, Q_B, Q_C ^b	3.3×10^4
10^4 WT-RNA	Q_A, Q_B, Q_C	4.7×10^3
10^4 WT-RNA + Q_A, Q_B, Q_C		2.5×10^4
	Viral stock RNA + Q_A, Q_B, Q_C ^b	2.1×10^4
Viral stock RNA	Q_A, Q_B, Q_C	2.0×10^3
Viral stock RNA + Q_A, Q_B, Q_C		1.5×10^4

a. Q_A , Q_B and Q_C were added at 10^2 , 10^3 and 10^4 RNA molecules, respectively.

b. Control quantifications with no nucleic acid re-isolation involved.

EXAMPLE 5:

The method according to the invention was compared to earlier described Q-NASBA protocols (Van Gemen et al.1993; Jurriaans et al., submitted, 1993) where only one Q-RNA is used, and, if a dynamic range of 5 logs is to be achieved,

at least 6 amplification reactions per clinical (wild-type) sample are necessary, i.e. one positive WT control without addition of internal standard and 5 reactions with an increasing amount ($10^2 - 10^6$) of internal standard RNA molecules.

The number of amplification reactions can be decreased with the method according to the invention where several distinguishable internal standards are spiked into one amplification.

In contrast, when only one Q-RNA is used (Q_A) in combination with enzyme labelled probes the initial WT-RNA concentration must be deduced from the ratio of the WT-signal to Q-signal using different concentrations Q-RNA in separate amplifications.

The two methods were compared using model systems and plasma samples of HIV-1 infected individuals.

The one-tube Q-NASBA using Q_A , Q_B and Q_C RNA was compared to the earlier described Q-NASBA protocol (Van Gemen et al., 1993) that uses 6 amplifications per quantification by analysing 0.1 ml plasma samples of 3 asymptomatic HIV-1 infected individuals. In the same experiment the difference between addition of Q_A , Q_B and Q_C before and after nucleic acid isolation was investigated again (table 3). In cases where the WT-RNA is quantitated by the one-tube Q-NASBA protocol the WT-RNA amount can be quantitated reliably to $1/10$ of the lowest Q-RNA amount used in the one tube quantification protocol.

Since the addition of Q_A , Q_B and Q_C (at 6×10^5 , 6×10^4 and 6×10^3 , respectively) is to 0.1 ml plasma when they are added to the lysis buffer, the lower reliable quantification limit is 6×10^2 RNA copies per 0.1 ml plasma. Addition of Q_A , Q_B and Q_C (at 10^4 , 10^3 and 10^2 , respectively) to the isolated nucleic acid gives a reliable lower quantification limit of 10^3 RNA copies per 0.1 ml plasma when 1 μ l plasma equivalents of nucleic acid are used for amplification. In both cases the WT-RNA

amount can be quantitated reliably to $1/10$ of the lowest Q-RNA amount used in the one-tube quantification protocol. The protocol using 6 amplifications per quantification with 10^2 as lowest internal standard Q-RNA amount has a lower reliable quantification limit of 10^4 RNA copies per 0.1 ml plasma when 1 μ l plasma equivalents of nucleic acid are used for amplification. The results indicate that in this particular experiment the efficiency of nucleic acid isolation varied between 100% (patient 1) and 50% (patient 2). Patient 3 was always below the reliable quantification limit, although this limit was most accurate (i.e. the lowest) for the one-tube Q-NASBA assay with addition of Q_A , Q_B and Q_C to the lysis buffer before nucleic acid isolation (table 3).

Table 3: Comparison of the one tube Q-NASBA with the quantification protocol using 6 amplifications per quantification on 1 ml plasma samples of 3 HIV-1 infected patients.

	A	B	C
Patient			
1	$<1.0 \times 10^4$	1.5×10^3	1.1×10^3
2	1.0×10^4	5.6×10^3	1.3×10^4
3	$<1.0 \times 10^4$	$<1.0 \times 10^3$	$<6.0 \times 10^2$

- A: Q-NASBA with 6 amplifications per quantification
 B: One-tube Q-NASBA; Q_A, B, C added after nucleic acid isolation.
 C: One-tube Q-NASBA; Q_A, B, C added before nucleic acid isolation.

EXAMPLE 6:

Finally we tested the reproducibility of the one-tube Q-NASBA with addition of Q_A , Q_B and Q_C to the lysis buffer before nucleic acid isolation using an *in vitro* cultured HIV-1 viral stock solution in which the amount of viral particles was quantitated by electronmicroscopy (Layne, S.P. et al., Virology. 189, 695-714, 1992). The HIV-1 viral stock containing 2.9×10^{10} viral particles per ml was diluted 10.000 times in water and Q_A , Q_B and Q_C (at 6×10^5 , 6×10^4 and 6×10^3 , respectively) were added to 100 μ l diluted viral stock solution, resulting in the measurement of 4.35×10^{10} RNA molecules per ml. Results are depicted in Table 4. The mean \pm SD of the RNA quantification was $10^{10.64 \pm 0.05}$, showing that the accuracy of the one-tube Q-NASBA is within 0.1 log when quantitating this *in vitro* cultured viral stock solution.

(The procedures followed are described in the Materials and Methods section of example 3).

From table 4 it can be seen that an accuracy of the assay within 0.1 log when quantitating HIV-1 RNA in an *in vitro* cultured viral stock solution can be achieved with the method according to the invention. This result enables reliable measurements of differences in HIV-1 RNA load of 0.4 logs and greater.

Table 4: Reproducibility of the one-tube Q-NASBA. The in vitro cultured HIV-1 viral stock contains $2.9 (\pm 1.6) \times 10^{10}$ viral particles per ml.

Nucleic acid isolation	RNA molecules per ml (mean \pm std)
1	$10^{10.69 \pm 0.16}$ a
1	$10^{10.53 \pm 0.03}$ a
1	$10^{10.61 \pm 0.03}$ a
2	$10^{10.64 \pm 0.05}$ a
2	$10^{10.65 \pm 0.01}$ a
2	$10^{10.68 \pm 0.02}$ a
3	$10^{10.64 \pm 0.05}$ b
4	$10^{10.67 \pm 0.06}$ b
5	$10^{10.65 \pm 0.07}$ b

Q_A , Q_B and Q_C (6×10^5 , 6×10^4 and 6×10^3 , respectively) were added to 100 μ l of a 10.000 dilution of the viral stock in water.

a. duplicate amplifications

b. triplicate amplifications

The mean \pm SD of all quantifications is $10^{10.64 \pm 0.05}$ (4.35×10^{10}) RNA molecules per ml.

CLAIMS:

1. Method for the quantification of analyte nucleic acid in a sample comprising the steps of:
 - adding to the sample different respective amounts of different nucleic acid constructs, each construct being distinguishable from the analyte nucleic acid and capable of being co-amplified with the analyte nucleic acid,
 - subjecting the sample to a nucleic acid amplification procedure, using amplification reagents capable of reacting with both the analyte nucleic acid and the nucleic acid constructs,
 - detecting the relative amounts of amplificates derived from analyte nucleic acid and each nucleic acid construct,
 - calculating the amount of analyte nucleic acid from said relative amounts.
2. Method according to claim 1, characterized in that the nucleic acid constructs have the same sequence as the analyte nucleic acid except for a uniquely distinguishable sequence in each nucleic acid construct.
3. Method according to claim 2, wherein the uniquely distinguishable sequence is a randomized sequence comprising about 20 nucleotides.
4. Method according to any of the preceding claims, characterized in that nucleic acid constructs are added in a range of amounts, differing from each other by a constant factor.
5. Method according to any of the preceding claims, characterized in that nucleic acid constructs are added in a range of amounts, differing from each other by a factor 10.

6. Method according to claim 1 wherein the sample is prepared by subjecting a known quantity of test-fluid to a nucleic acid isolation procedure, characterized in that the the nucleic acid constructs are added to the test-fluid prior to subjecting said test-fluid to the nucleic acid isolation procedure.
7. Method according to claim 6, characterized in that a known quantity of an isolation control sequence is added to the sample after the test-fluid has been subjected to an nucleic acid isolation procedure.
8. Method according to any of claims 1-7, characterized in that the sample is split in more than one reaction volumes to each of which nucleic acid constructs are added in different ranges of amounts.
9. Method according to claim 8, characterized in that to each reaction volume the same nucleic acid constructs are added.
10. Method according to claim 8 or 9, characterized in that the ranges of amounts do not overlap.
11. Method for the quantification of analyte nucleic acid in more than one sample wherein each sample is subjected to the method of claim 1, wherein the same nucleic acid constructs are used in all samples and the amounts of each particular nucleic acid construct are varied.
12. Method according to claim 11, characterized in that in each sample a zero amount of one of the nucleic acid constructs is used.
13. Method for the quantification of analyte nucleic acid in a panel of test fluids comprising the steps of:
 - diluting a quantity of said test fluids by a known factor and taking samples from said diluted test

fluids containing an amount of analyte nucleic acid expected to be within a certain range,

- subjecting the samples to the method of claim 1 wherein the amounts of nucleic acid constructs added to the samples are within said range.

14. Method according to claim 13, comprising the additional steps of:
 - identifying samples containing an amount of analyte nucleic acid lower than said range of amounts,
 - subjecting the test-fluids from which said samples originate to the method of claim 13, wherein said quantities are diluted by a lower factor.
15. Method according to claim 13, comprising the additional steps of:
 - identifying samples containing an amount of analyte nucleic acid higher than said range of amounts,
 - subjecting the test-fluids from which said samples originate to the method of claim 13, wherein said quantities are diluted by a higher factor.
16. Method for the quantification of analyte nucleic acid in a sample comprising the steps of:
 - adding to a sample a quantity of a nucleic acid construct said quantity expected to be in the same range as the expected amount of analyte nucleic acid present in said sample.
 - subjecting the sample to a nucleic acid amplification procedure, using amplification reagents capable of reacting with both the analyte nucleic acid and the nucleic acid construct,
 - detecting the relative amount of amplicates derived from analyte nucleic acid and the nucleic acid construct,
 - estimating the amount of analyte nucleic acid present in said sample from said relative amount
 - subjecting a second sample from the same test fluid to the method of claim 1, wherein the amounts of

nucleic acid constructs added are within the same range as the estimated amount of analyte nucleic acid present in the first sample.

17. Test kit for the quantification of nucleic acid in a sample according to the method of claim 1, comprising different nucleic acid constructs, amplification reagents capable of reacting with the nucleic acid constructs and the analyte nucleic acid, a detection probe for the amplified analyte nucleic acid and different detection probes specific for the different amplified nucleic acid constructs respectively.

FIGURE 1A

Theoretical co-amplification of RNA
Sample: 150 Wild-type copies

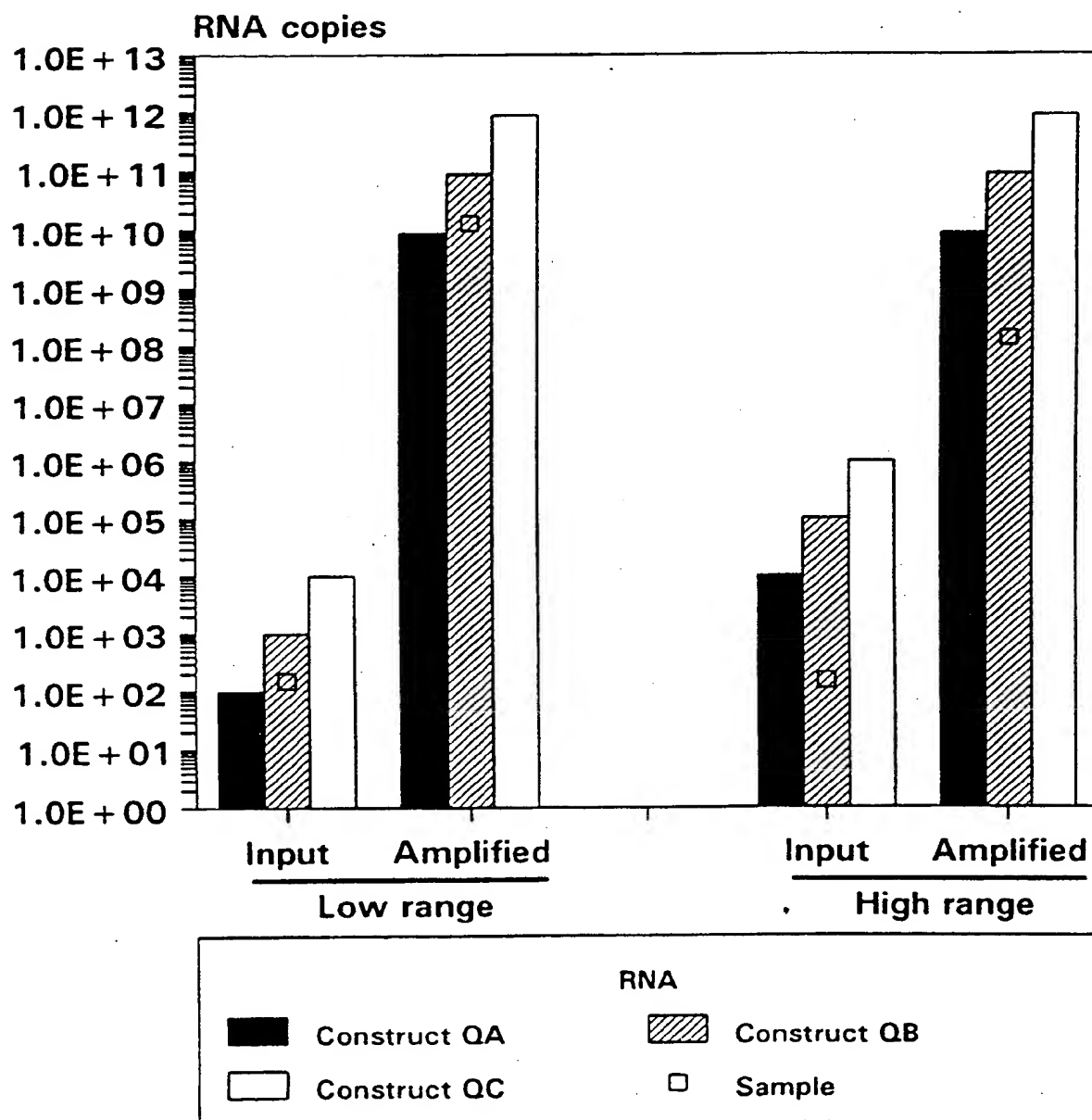


FIGURE 1B

Detection Assay

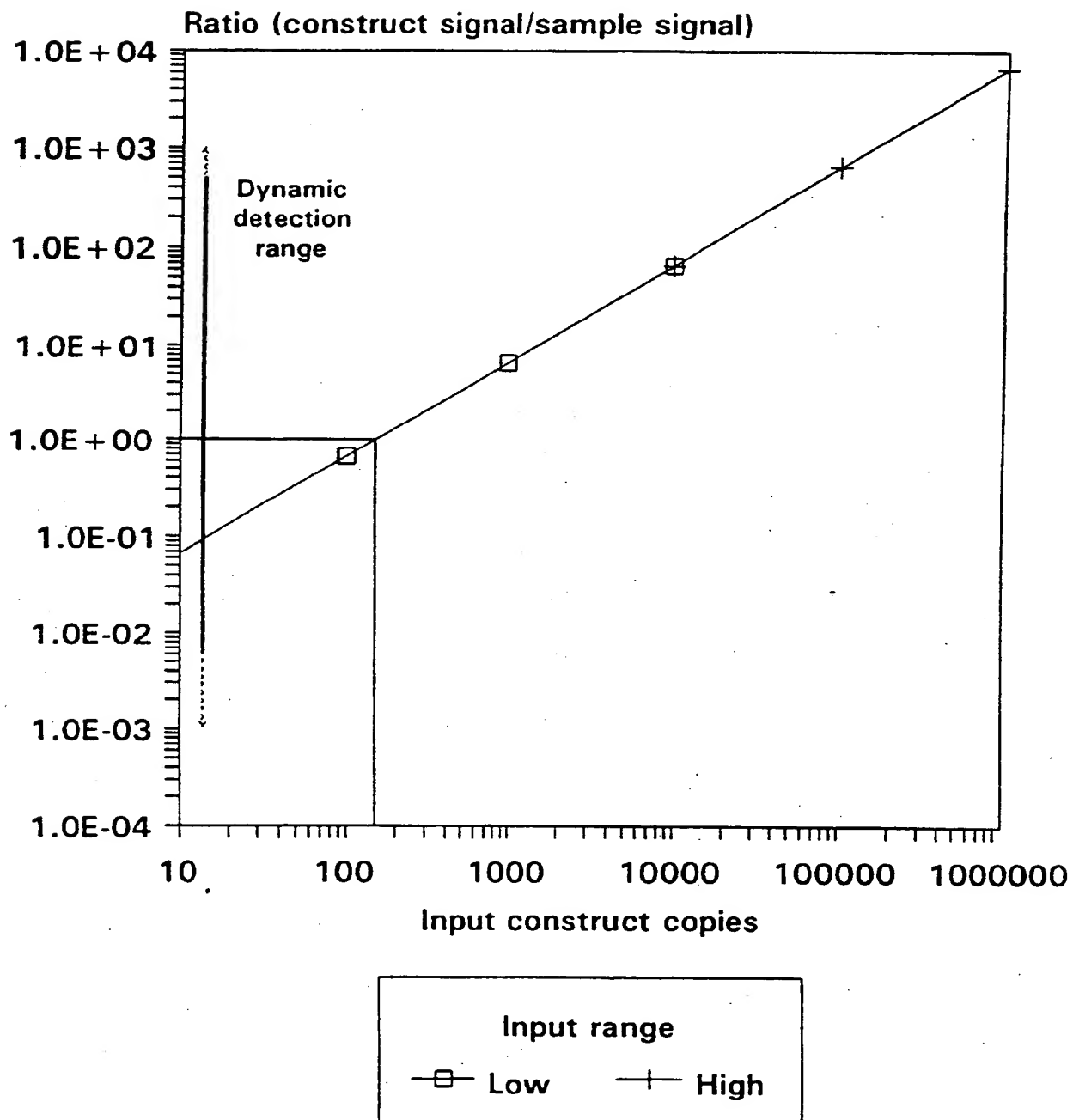


FIGURE 2A

Theoretical co-amplification of RNA
Sample: 2000 Wild-type copies

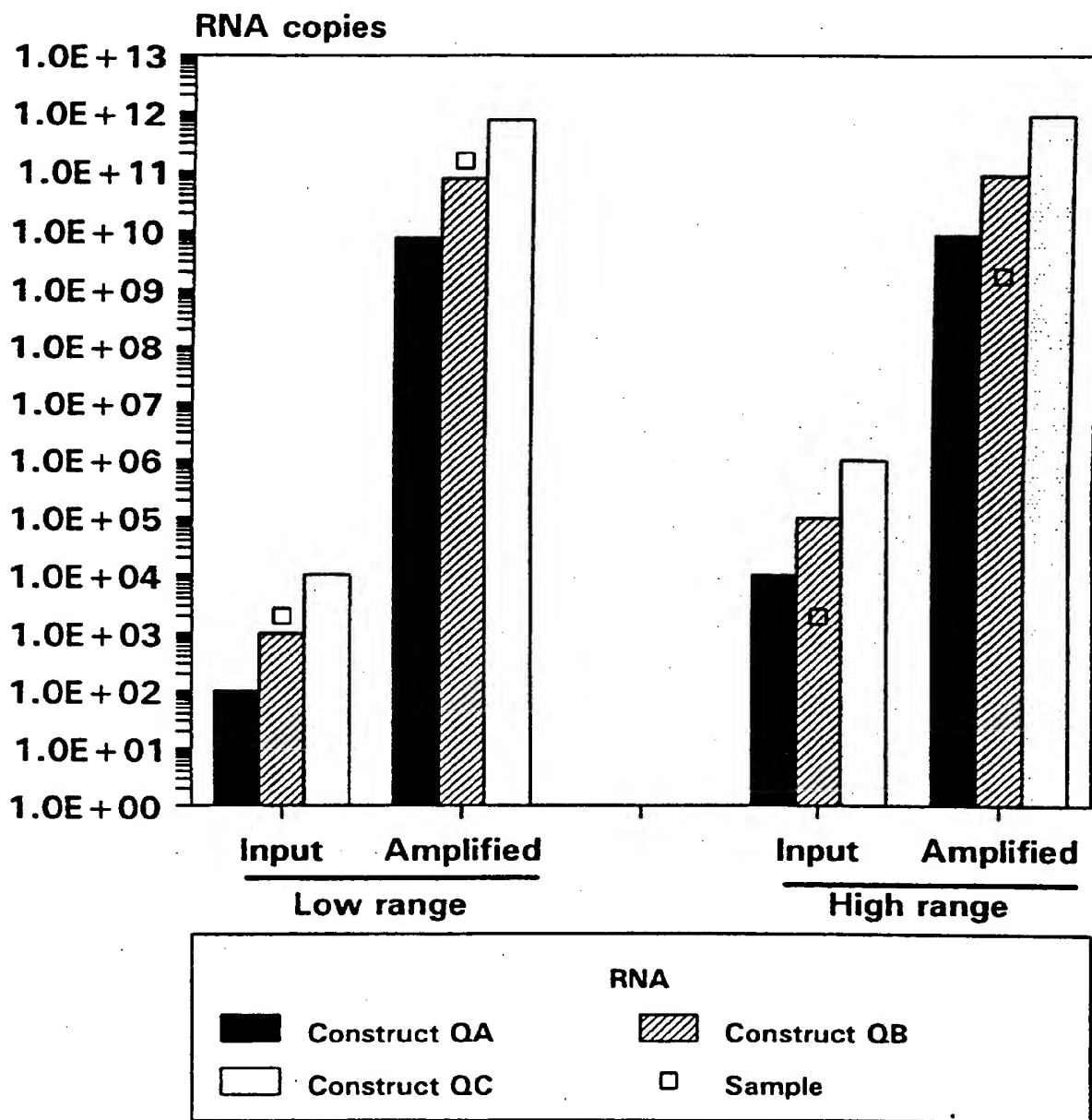


FIGURE 2B

Detection Assay

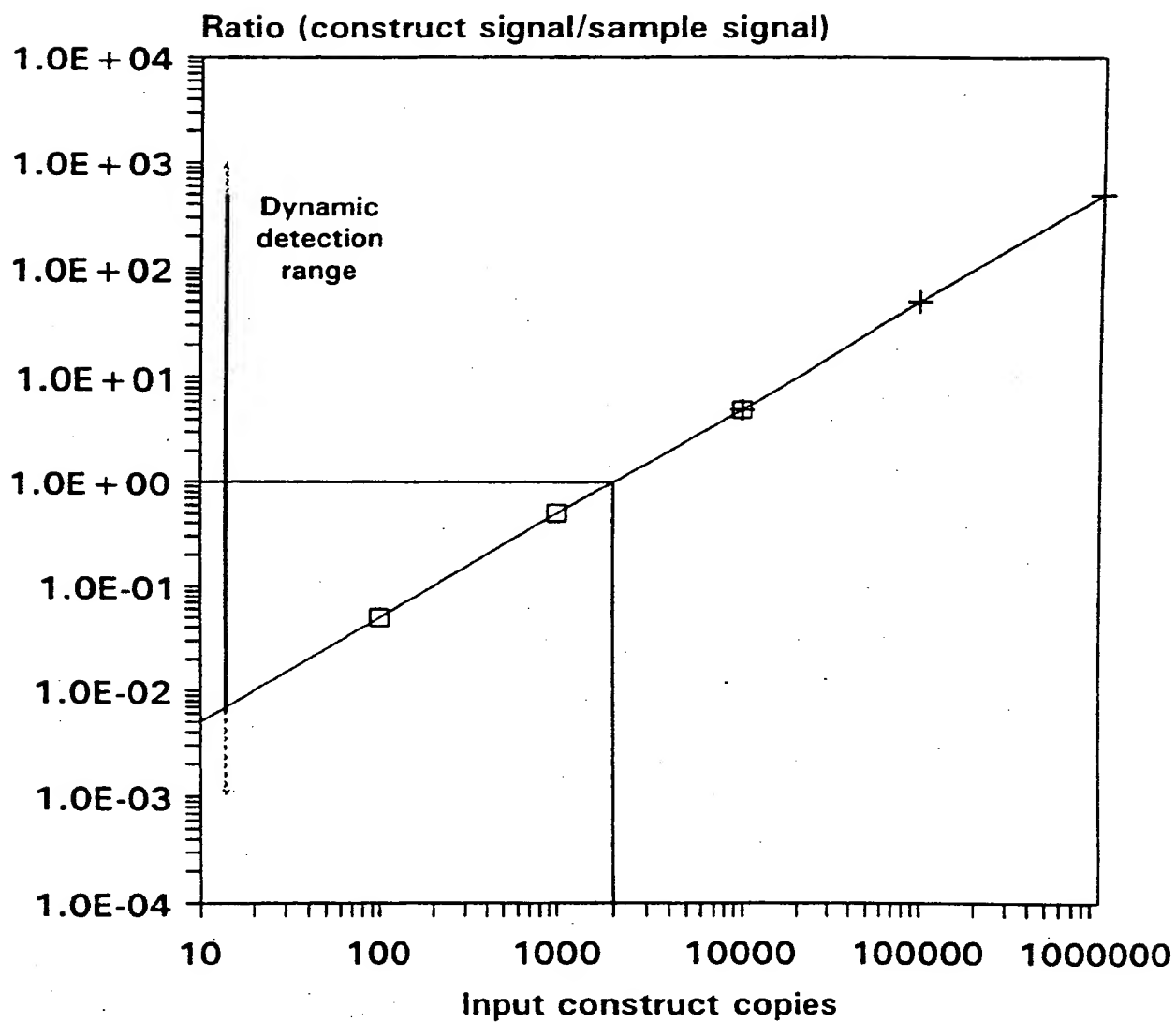


FIGURE 3A

Theoretical co-amplification of RNA
Sample: 300000 Wild-type copies

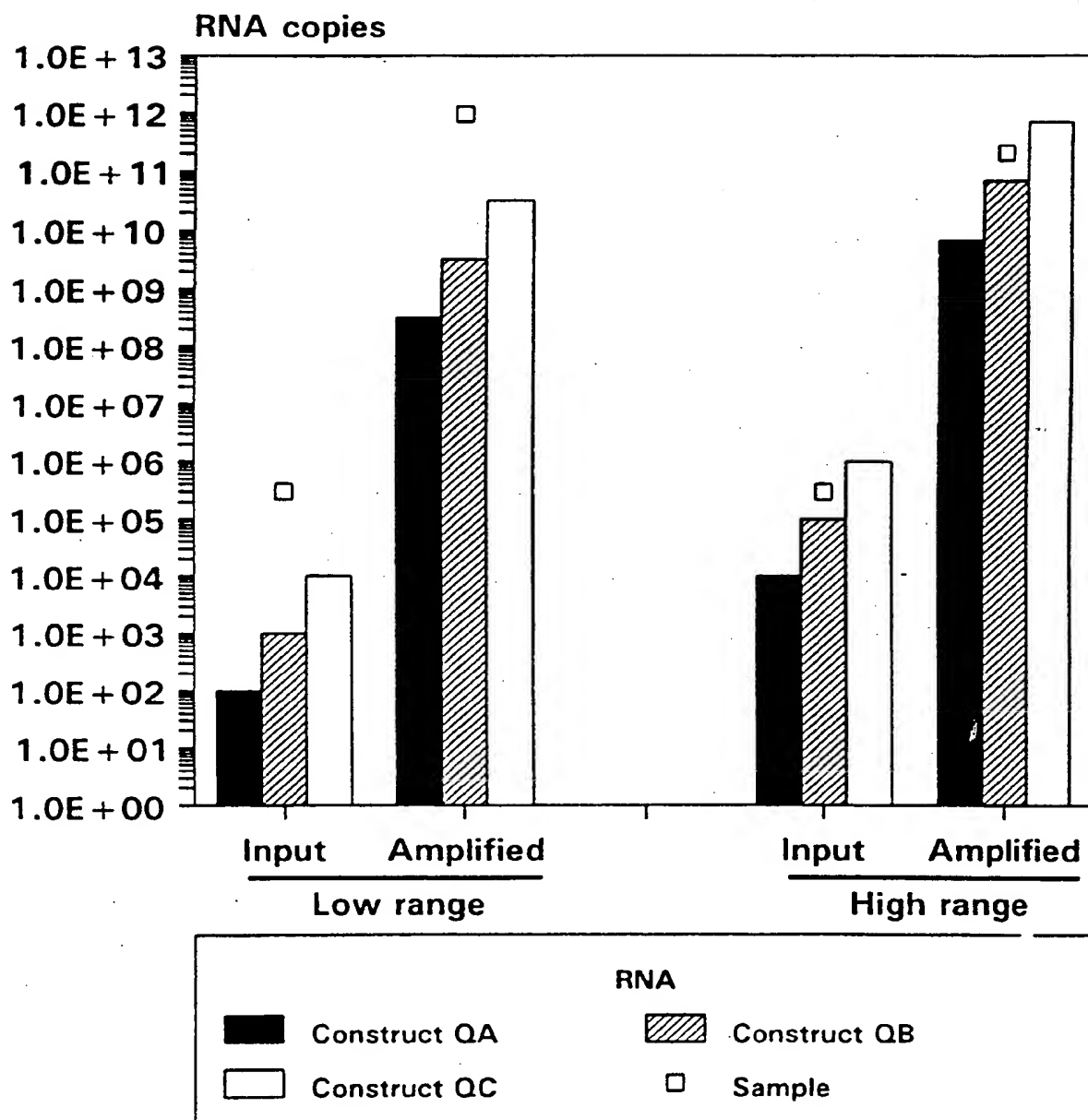


FIGURE 3B

Detection Assay

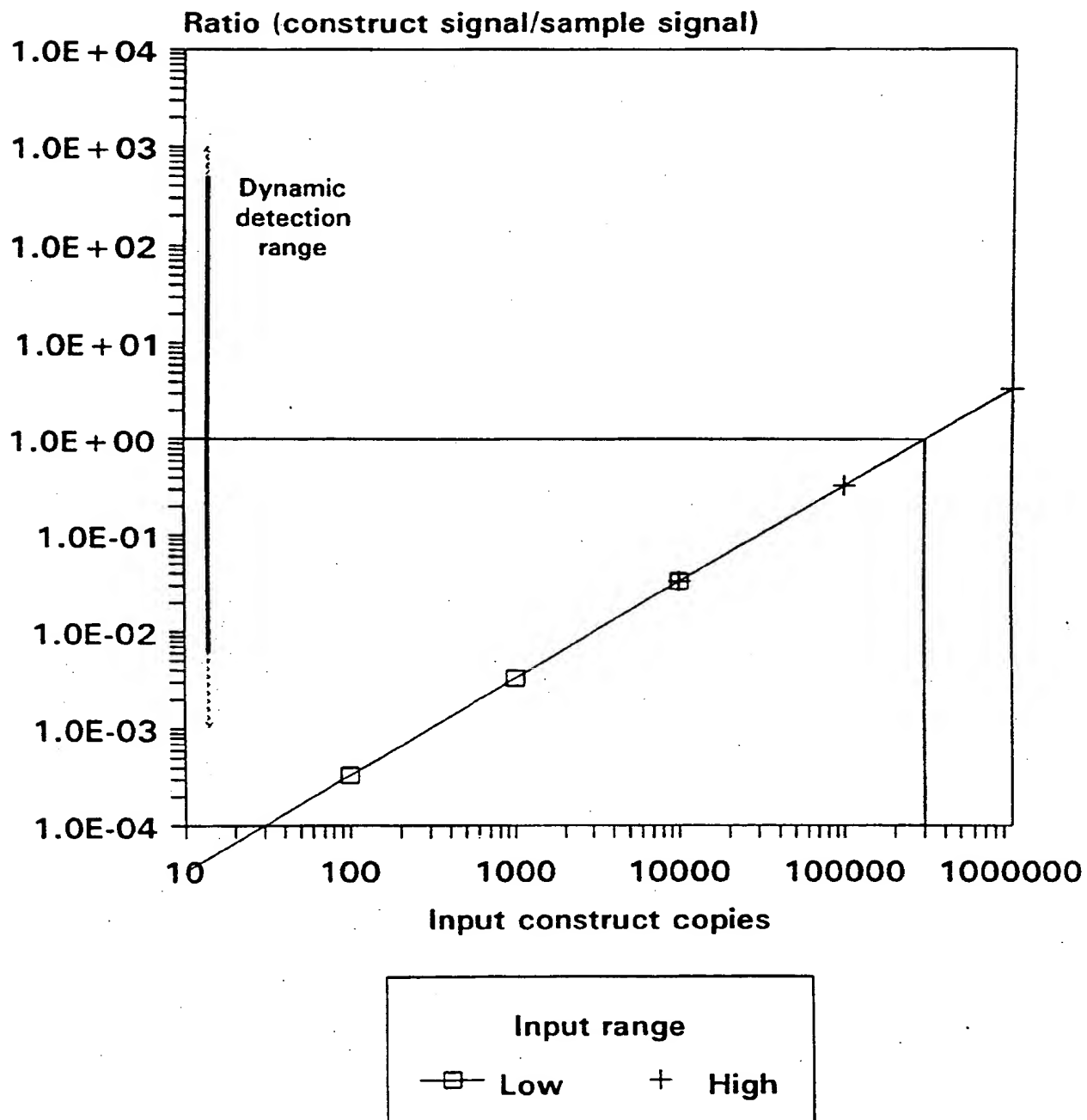


FIGURE 4A

**Theoretical co-amplification of RNA
Using two instead of three constructs
Sample: 5000 Wild-type copies**

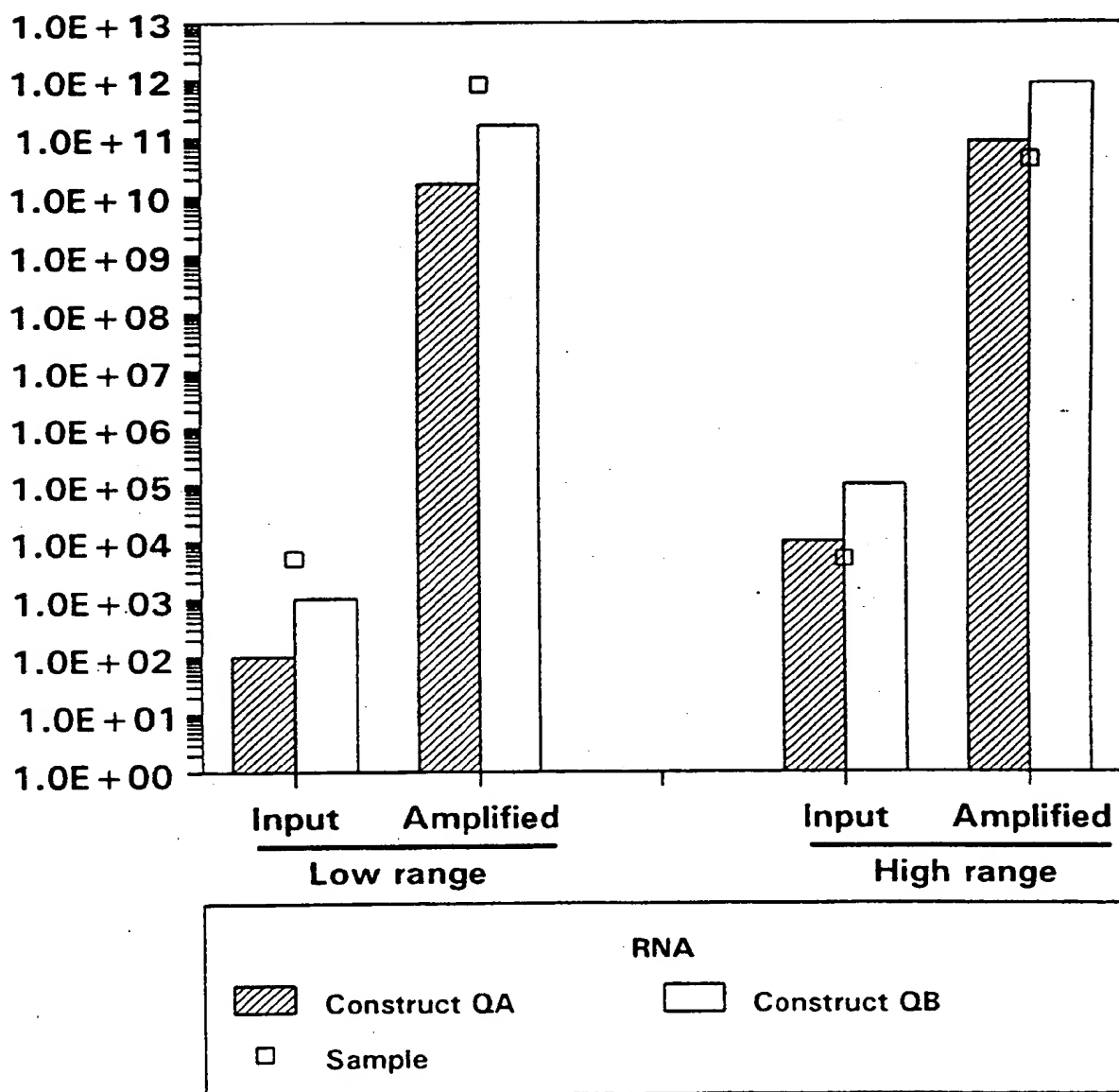


FIGURE 4B

Detection Assay

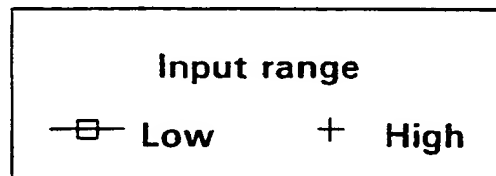
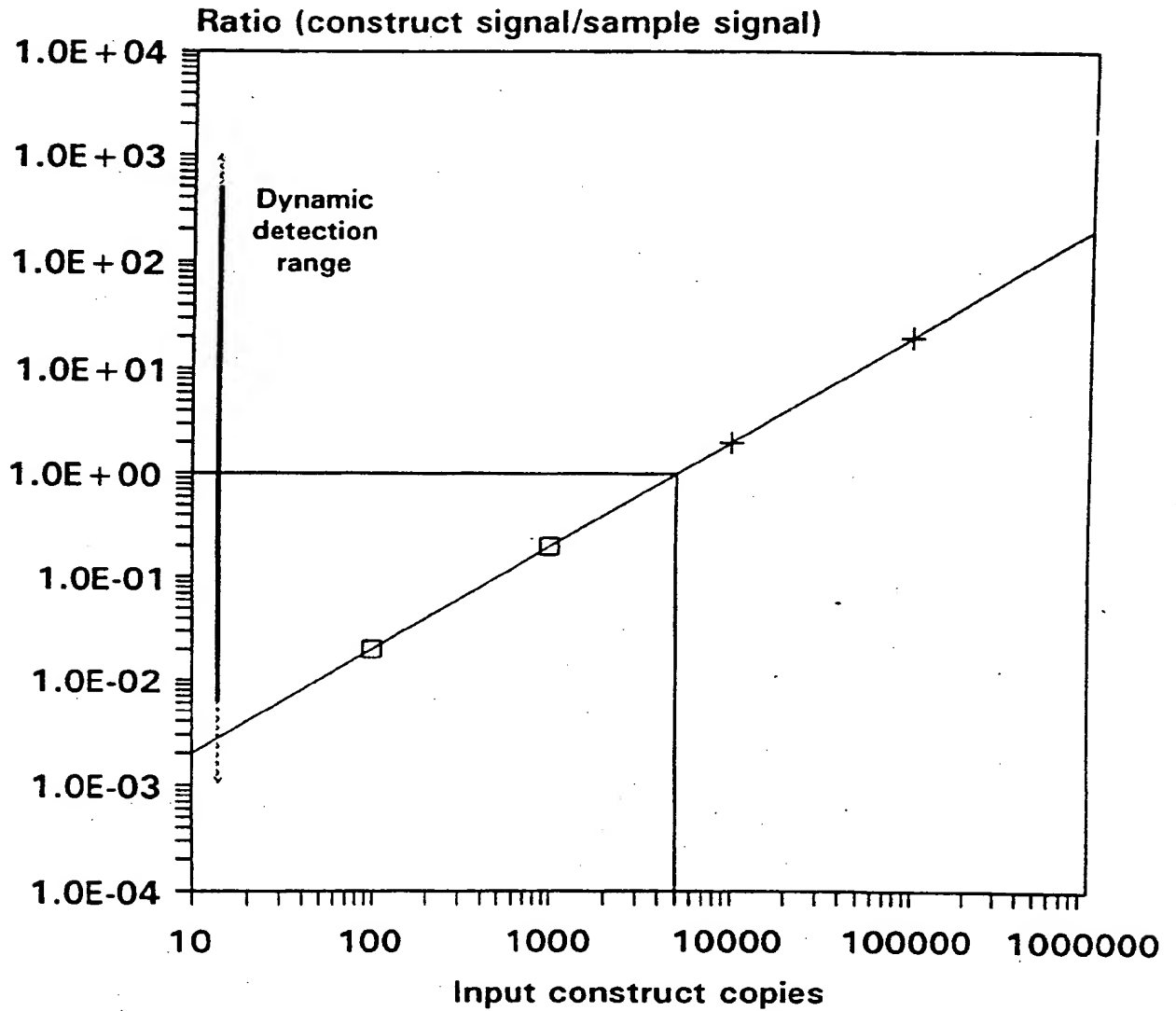
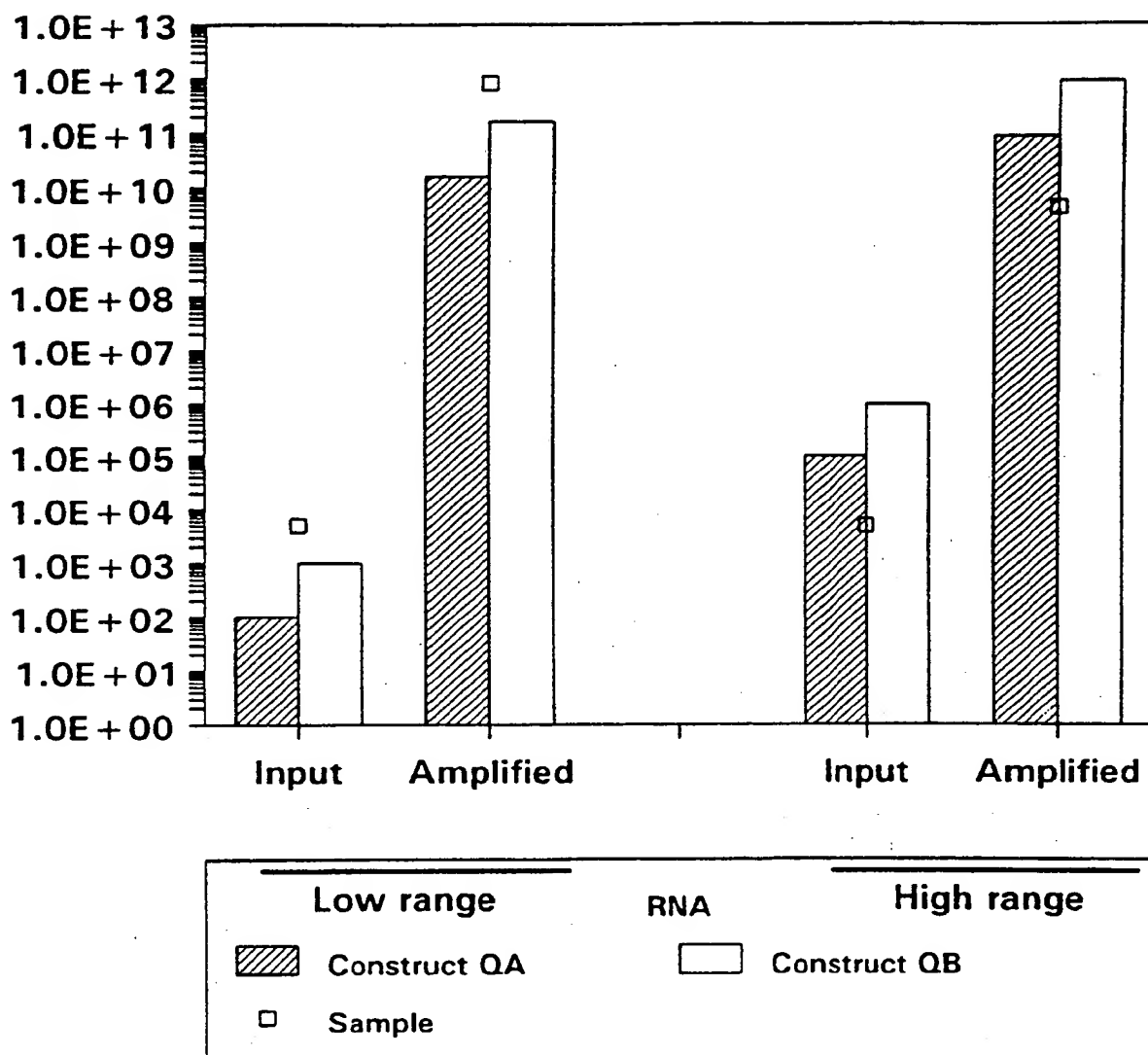


FIGURE 5A

**Theoretical co-amplification of RNA
Using two instead of three constructs
Sample: 5000 Wild-type copies**



**A gap was used to expand the
dynamic range**

FIGURE 5B

Detection Assay

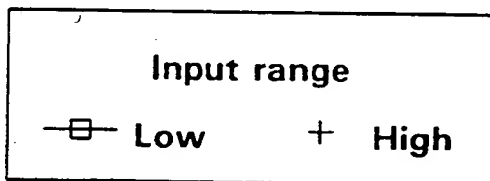
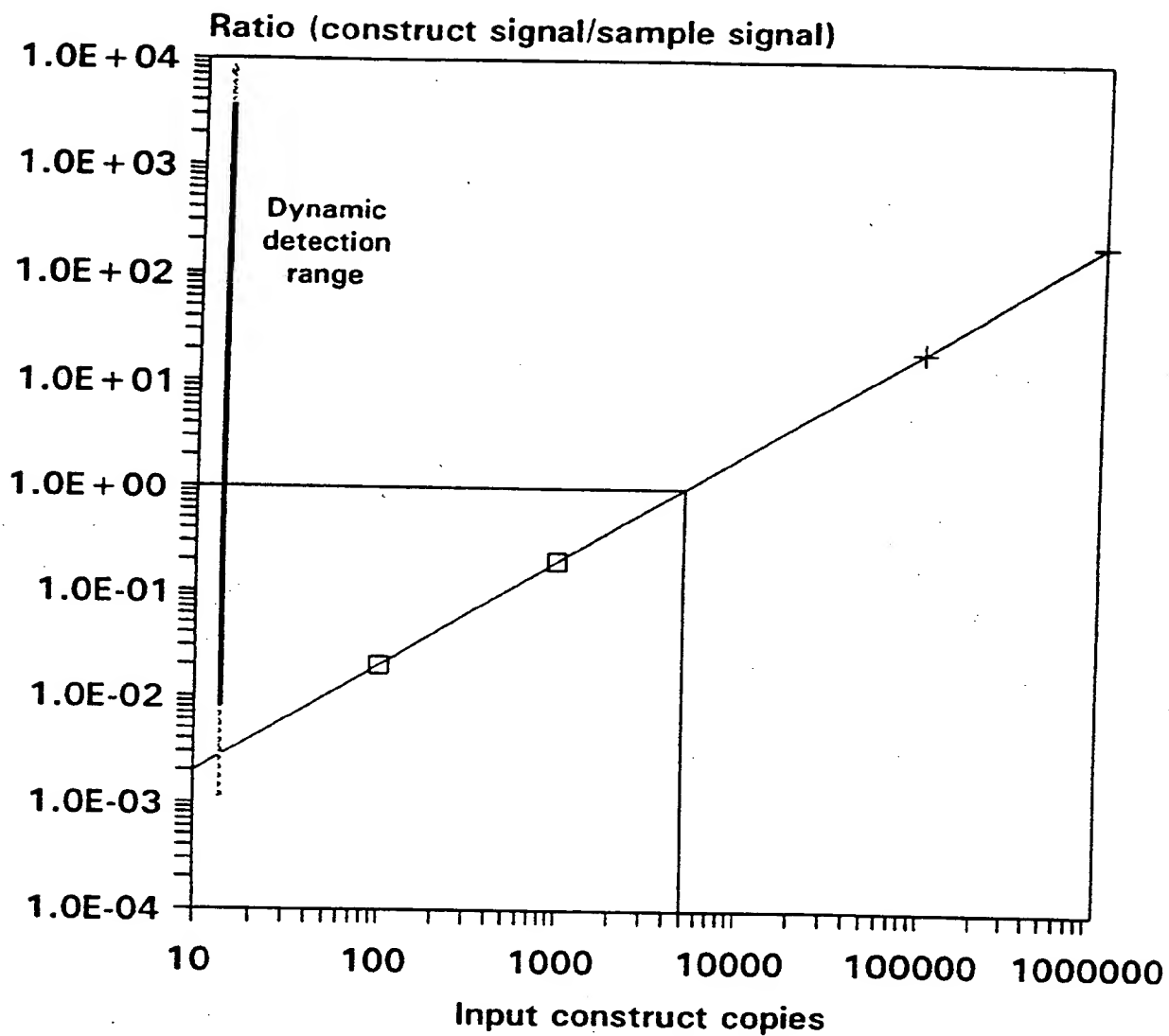


FIGURE 6

Dynamic range Q-NASBA

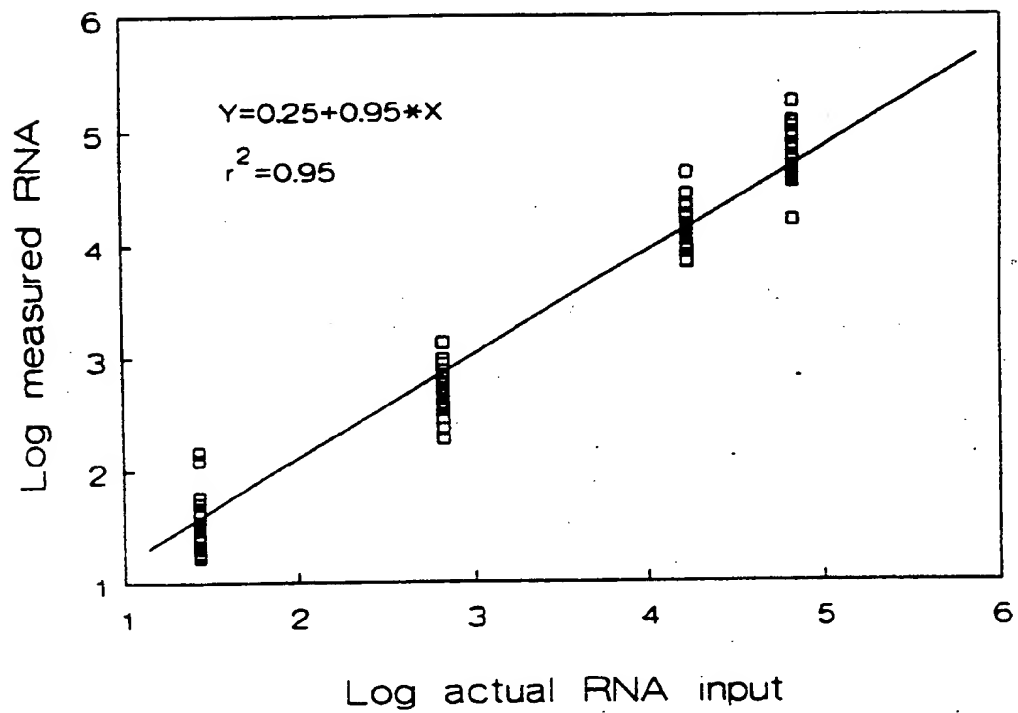
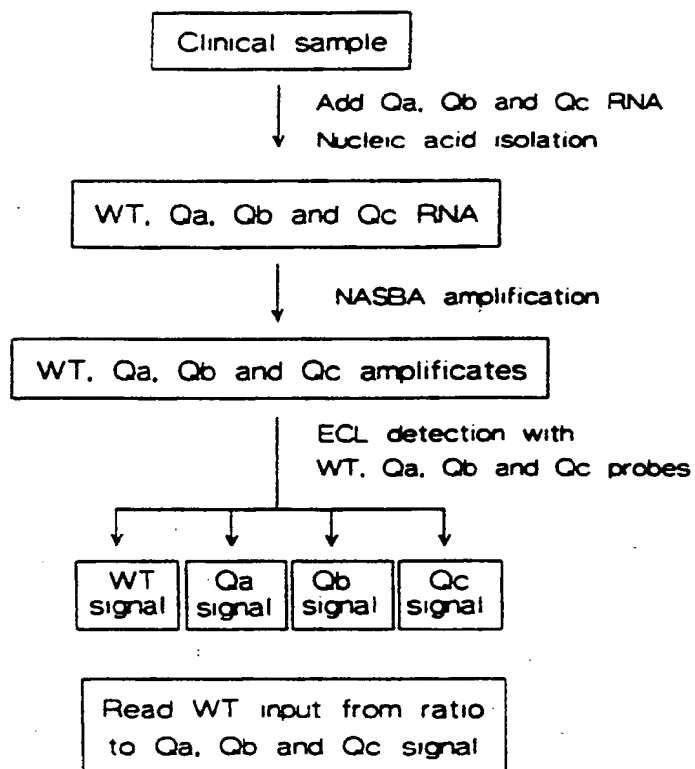


FIGURE 7



INTERNATIONAL SEARCH REPORT

International Application No
PCT/EP 94/02295

A. CLASSIFICATION OF SUBJECT MATTER
IPC 6 C12Q1/68 C12Q1/70

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 6 C12Q

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	EP,A,0 525 882 (AKZO) 3 February 1993 cited in the application see the whole document ---	1-3
Y	JOURNAL OF VIROLOGICAL METHODS, vol.43, July 1993, AMSTERDAM NL pages 177 - 188 GEMEN ET AL. cited in the application see the whole document ---	1-3
Y	EP,A,0 303 459 (PRESIDENT AND FELLOWS OF HARVARD COLLEGE) 15 February 1989 see the whole document ---	1-3
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☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

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- *O* document referring to an oral disclosure, use, exhibition or other means
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Date of the actual completion of the international search

3 November 1994

Date of mailing of the international search report

28. 11. 94

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Authorized officer

Molina Galan, E

INTERNATIONAL SEARCH REPORT

International Application No
PCT/EP 94/02295

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO,A,93 10257 (INSTITUT NATIONAL DE LA SANTÉ ET DE LA RECHERCHE MEDICALE) 27 May 1993 see page 44; claims ---	1
P,A	ANALYTICAL BIOCHEMISTRY, vol.213, September 1993, NEW YORK US pages 277 - 284 APOSTOLAKOS ET AL. -----	

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/EP 94/02295

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
EP-A-0525882	03-02-93	AU-A- 2071892 JP-A- 5219999	11-03-93 31-08-93
EP-A-0303459	15-02-89	US-A- 4942124 JP-A- 1137982 US-A- 5149625	17-07-90 30-05-89 22-09-92
WO-A-9310257	27-05-93	FR-A- 2683827 CA-A- 2123680 EP-A- 0613503	21-05-93 27-05-93 07-09-94

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